

Article

# The Vitamin k<sub>2</sub> improves endothelial progenitor cells vascular repair in rats' dyslipidaemia: an experimental study

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**Abstract.** *Background:* Vitamin k<sub>2</sub> (menaquinone-7) was reported to possess a vascular protective action against atherogenesis through reduction of vascular calcification.

*Objectives:* The present study investigated whether the augmentation of endothelial progenitor cells' (EPCs) reparative capacity could be an underlying mechanism behind vitamin k<sub>2</sub> vasoprotective action, in a rat model of dyslipidaemia.

*Methods:* Forty-five Wistar rats were randomly assigned to normal control (15-rat) or dyslipidaemic rats (30-rat) fed on laboratory show or high fat diet (HFD), respectively. Dyslipidaemic rats were further assigned to receive either vehicle or vitamin k<sub>2</sub> (30 mg/kg) 5 days a week, orally for 8 weeks. At the end of the study, lipid profile was assessed. Thoracic aortae were dissected for histopathological examination, immunostaining for detection of EPCs markers; CD133 and vascular endothelial growth factor receptor-2 (VEGFR-2), and  $\beta$ -catenin expression, and for expression of NADPH oxidase 4 (NOX4). Vascular function was assessed biologically in-vitro. *Results:* Vitamin k<sub>2</sub> supplementation conferred an endothelial protection and anti-atherogenic potential when compared to vehicle treated HFD fed rats evidenced by improved lipid profile; serum TG (mg/dl): 109.4  $\pm$  16.56 versus 153.60  $\pm$  9.88, serum LDL: 53.56  $\pm$  12.76

versus  $89.00 \pm 9.80$  and atherogenic index of  $0.52 \pm 0.12$  versus  $0.69 \pm 0.05$ , respectively. Moreover, The significant increase in EPCs numbers induced by vitamin k2 treatment in comparison to vehicle treated HFD fed rats;  $33.50 \pm 4.73$  versus  $22.30 \pm 3.30$ , respectively, appears to play a pivotal role in the vaso-protective action of vitamin K<sub>2</sub> that could be mediated by interplay between vascular Wnt/ $\beta$ -catenin signalling and NOX4 expression.

*Conclusion:* 8-week treatment with vitamin k<sub>2</sub> increases EPCs count as well as confers anti-atherogenic potential in rats with dyslipidemia and whether continued vitamin k<sub>2</sub> supplementation encounters sustained EPCs endothelial regeneration warrants further investigation.

**Keywords:** endothelial dysfunction, atherogenesis, CD133, VEGFR-2, NOX4

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## Introduction

Endothelial dysfunction (ED) is a hallmark of cardiovascular diseases and is considered the earliest event in pathogenesis of atherosclerosis. To establish endothelial homeostasis, endogenous repair mechanisms are operating in parallel to the endothelial constant insult. Endothelial progenitor cells (EPCs) play a vital role in vascular regeneration after injury and inhibition of atherosclerotic lesion progression. EPCs contribute to the pro-angiogenesis in ischemic tissue and vascular regeneration and remodelling.(1)

Dyslipidaemia, as a risk factor, switches the endothelium quiescent state into activated one which alters endothelial cell signalling leading ultimately to ED.(2) Accumulation of the lipid-containing lipoproteins in the intima activates the endothelium and initiates an atherosclerotic process and cardiovascular events.(3, 4) In dyslipidaemia, EPCs profile is altered and their circulating numbers are reduced which collaborate into the induction of ED,(5) though the underlying mechanisms are still not clear.

Oxidative stress could be one of these mechanisms. In fact, NADPH oxidases (NOXs) are major sources of endothelial reactive oxygen species (ROS). Although ROS are mainly involved in the physiological vascular homeostasis, excess ROS is pathological and results in oxidative stress leading to ED and vascular disease progression.(6) The endothelial physiological homeostasis is thought to be maintained by the constitutively expressed NOX4 (7).

The EPCs reparative function process remains obscure and needs further investigations. One of the master regulators of EPCs enrolment and differentiation is the canonical Wnt signalling pathway.(8) An interaction between Wnt and ROS-producing NOX enzymes has been postulated; however the individual role of NOX4 is not clear yet.(9) NOX1 and NOX4 were demonstrated to activate the p38 MAPK pathway,(10) which phosphorylates and inhibits glycogen synthase kinase (GSK-3 $\beta$ ), thereby maintaining Wnt/ $\beta$ -catenin in a stabilized state during cellular differentiation.

Thus, interplay could be raised between NOX4 expression and Wnt/ $\beta$ -catenin signalling in vascular repair mechanisms. Being a marker for ED, EPCs became an attractive target for vascular repair. Therefore, it is mandatory to explore more effective EPC-based therapies.(11)

In this context, vitamin k<sub>2</sub>, as a nutritional supplement, was of concern. Vitamin K<sub>2</sub> (Menaquinones) is obtained from meats, a variety of cheeses, and eggs. Besides reduction of coronary calcification and atherogenesis, vitamin K<sub>2</sub> enrolment in maintenance of endothelial cells' survival by rescuing it from apoptosis has been raised.<sup>(12)</sup> In the present study, whether vitamin K<sub>2</sub> can improve the detrimental effect of dyslipidemia on EPCs reparative capacity is investigated in a rat model of dyslipidaemia and its possible relation to the expression levels of NOX4 and  $\beta$ -catenin proteins is further unravelled.

## Materials and Methods

### *Animals*

The study was conducted on male Wistar albino rats weighing 190-230g. Animals were housed under standard conditions with food and water ad libitum. All animals' procedures and treatment were conducted in accordance with the Research Ethics Committee - Faculty of Medicine, Alexandria University, in compliance with the "Public Health Service (PHS) Policy on Humane Care and Use of Laboratory Animals Policy".

### *Experimental procedure*

After acclimatization, 45 rats were randomly assigned into 2 main groups according to the fed diet for 8 weeks. Normal control rats (n=15) were fed on laboratory chow (1.29 KJ as fat) and a high-fat diet (HFD, 7.74 KJ/g as fat) was served to the dyslipidaemia-induced group of rats (n=30)<sup>(13)</sup> The dyslipidaemia-induced group of rats was then randomly subdivided into two subgroups (15-rat each) based on the assigned oral treatment that started with the dietary regimen for 8 weeks; vitamin K<sub>2</sub> (Sigma-Aldrich - 30 mg/kg/day, 5 days a week),<sup>(14)</sup> and vehicle (Gum acacia 2%) treated subgroup. Throughout the 8 weeks, rats were closely observed for any sign of humane endpoints. At the end of the study after an overnight fasting, rats were anesthetized by intraperitoneal thiopental sodium (50 mg/kg) and blood samples were collected by cardiac puncture on EDTA-containing tube and stored at 4 °C for lipid profile estimation. Then, animals were sacrificed by an overdose of anaesthesia. Multiple arterial sections from thoracic aortae were gently dissected and put in fresh Krebs solution for *in-vitro* isometric tension studies. Other aortic sections were frozen at - 80°C for later biochemical measurement or preserved in 10% formol saline for histopathological and immunostaining assessments.

### *Serum lipids measurements*

Diagnostic kits for the lipid profile; Total cholesterol, High-density lipoprotein cholesterol, and triglycerides (TC, HDL-C, TG, mmol/L) (with the exception of Low density lipoprotein-cholesterol, LDL-C) were purchased from BioSystems (S.A Costa Brava, Spain). The assays were performed according to the manufacturer's instruction. The LDL-C and atherogenic index of plasma (AIP) were calculated using Friedewald equation and logarithmic transformation of the ratio of TG to HDL-C, respectively.<sup>(15, 16)</sup>

#### *NOX4 gene expression by qPCR*

The abdominal aortic tissues were homogenised and total serum RNA was isolated using miRNeasy Mini Kit (QIAGEN), according to the manufacturer's protocol. The concentration of total RNA samples was quantified by a Nanodrop 2000 (Nanodrop, USA). The TaqMan MicroRNA Reverse Transcription (RT) Kit (Applied Biosystems, USA) was used for the RT reaction. The PCR was performed using a reaction volume of 20  $\mu$ L. The plate was prepared and ABI prism 7900 sequence detection system (Ambion, USA) was used for amplification and detection by qRT-PCR. Amplification of both target and housekeeping genes ( $\beta$ -actin) was performed using TaqMan Universal PCR master mix (Applied Biosystems), 25 ng cDNA, and the pre-designed probe and primer sets for rat specific NOX4 genes (TaqMan Gene Expression Assays, Applied Biosystems).(17)

#### *Histological sections and immunostaining*

##### *Histological examination*

After processing into paraffin, thoracic aortic sections were sliced into 5  $\mu$ m transverse sections from each vessel and were stained with haematoxylin and eosin (H&E) stain and examined under light microscope for diagnosing and scoring of atherosclerotic changes according to the AHA classification of human atherosclerotic lesions.(18)

##### *Immunostaining*

Immunohistochemistry (IHC) of aortic sections for EPCs surface markers CD133 and VEGF receptor-2, and for  $\beta$ -catenin was performed using prediluted (1:100 dilution) primary antibodies against CD133 (clone AC133; Miltenyi Biotec, Bergisch Gladbach, Germany); VEGF receptor-2 (clone KDR-2; Sigma-Aldrich, Missouri, USA), and  $\beta$ -catenin (Santa Cruz sc-7963). Immunoreactivity was developed using the streptavidin-biotin-immunoenzymatic antigen detection system (Neo Markers, Fremont, USA), which was performed according to manufacturer's protocol. The bound antibodies were detected using Ultra Vision Detection System Anti-Polyvalent, HRP/DAB (Dako REAL, EnVision™ Detection System, Denmark). Positive immunostaining was defined as a cytoplasmic and/or membranous staining of cells. Positive and negative controls were included in all runs. Staining density was semi-quantitatively assessed for each antibody separately and was defined as the percentage of positively stained endothelial cells to the total number of endothelial cells in ten randomly selected high-power fields (HPFs  $\times$ 400).(19) The combined EPCs surface markers (CD133+/VEGF receptor-2++) reflect a subpopulation of EPCs with vaso-regenerative functions.(20)

##### *Statistical analysis*

One-way Analysis of Variance (ANOVA) followed by Tukey as a post-hoc test for multiple comparisons or Kruskal-Wallis test followed by Dunnett's multiple comparison for mean ranks between groups were used for parametric and non-parametric data, respectively. Data analysis was done using Statistical package: MATLAB Statistical toolbox (Matrices Laboratory

software-Math Works ®, USA) and data were expressed as means ± SD or medians ± interquartile range. Statistical significance was set at  $p < 0.05$ .

## Results

### *Effect of vitamin K<sub>2</sub> on serum lipid profile*

Administration of HFD for 8 weeks exhibited a significant increase in atherogenic serum lipids and a significant decrease in serum HDL-C with a 50% higher atherogenic index compared to the normal control (Table 1). Treatment with vitamin k<sub>2</sub> significantly improved lipid profile, yet its induced increase in serum HDL-C was not significant versus the vehicle treated HFD rats with an atherogenic index of about 20% lower.

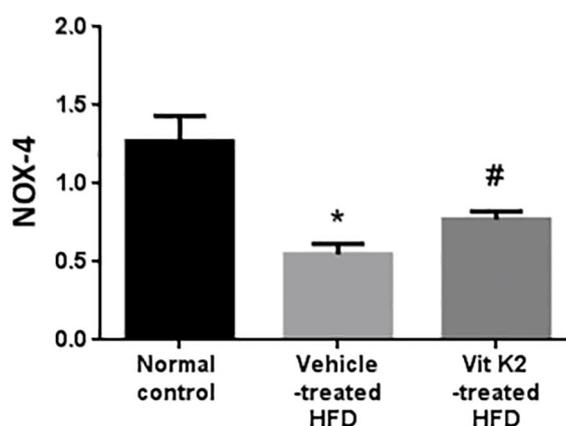
**Table 1: Effect of vitamin k<sub>2</sub> on serum lipid profile in HFD fed rats**

Lipid profile	Normal control	Vehicle-treated HFD	Vitamin k <sub>2</sub> - treated HFD
TC	115 ± 6.93	152± 8.37*	117.4± 8.53 <sup>#</sup>
TGs	104.40 ± 13.17	153.6± 9.88*	109.4± 16.56 <sup>#</sup>
LDL-C	45.22 ± 7.13	89 ± 9.80*	53.56 ± 12.76 <sup>#</sup>
HDL-C	50.00 ± 5.52	31.60± 3.95*	33.56 ± 5.92
Atherogenic index	0.32 ± 0.04	0.69±0.05*	0.52± 0.12 <sup>#</sup>

TC; total cholesterol, TGs; triglycerides, LDL-C; low-density lipoprotein cholesterol, HDL-C; high-density lipoprotein cholesterol, HFD; high fat diet. Data are expressed as means ± S.D. \*: significance difference versus normal control, <sup>#</sup>: significance difference versus vehicle-treated HFD.

### *Effect of vitamin K<sub>2</sub> on NOX4 gene expression*

Expression of NOX4 in aorta was significantly reduced in vehicle-treated HFD fed rats versus normal control, whereas, its expression was significantly increased after 8-week treatment with vitamin k<sub>2</sub> (Figure 1).



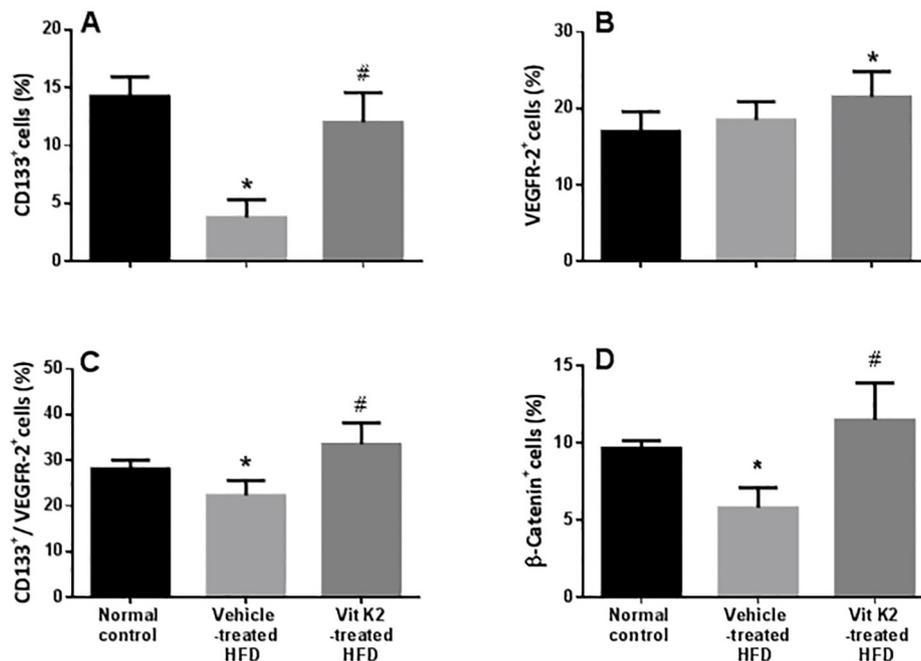
**Figure 1: Effect of vitamin k<sub>2</sub> on aortic NOX-4 expression.** Data are expressed as means ± S.D. \*: significant difference versus normal control, <sup>#</sup>: significant difference versus vehicle-treated HFD.

*Effect of vitamin k<sub>2</sub> on CD133/VEGFR2 expression*

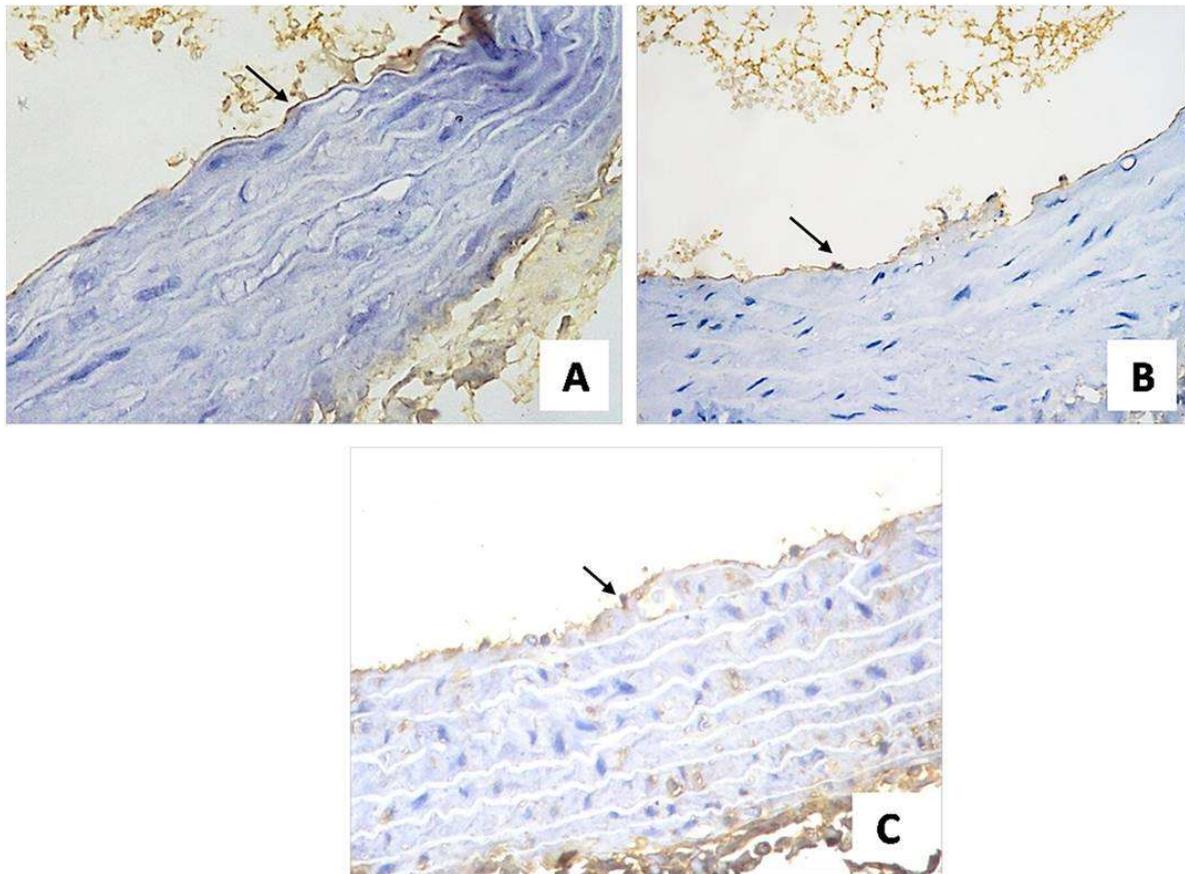
A significant decrease in percentage of CD133<sup>+</sup> cells was displayed in the endothelial layer obtained from vehicle-treated HFD rats versus normal control. Conversely, vitamin k<sub>2</sub> treatment resulted in a significant increase in CD133<sup>+</sup> cells percent (**Figure 2 A and Figure 3 A-C**). Regarding the percentage of VEGFR2<sup>+</sup> cells, it was non-significantly increased in vehicle-treated HFD rats versus normal control, whereas, vitamin k<sub>2</sub>-treated HFD rats showed a significant increase only versus normal control (**Figure 2 B, and Figure 4 A-C**). However, the percentage of combined EPCs surface markers (CD133/VEGFR2) was significantly decreased in vehicle-treated HFD rats versus normal control and it was significantly increased with vitamin k<sub>2</sub> treatment compared to vehicle-treated HFD rats (**Figure 2 C**).

*Effect of vitamin K<sub>2</sub> on  $\beta$ -catenin<sup>+</sup> cells*

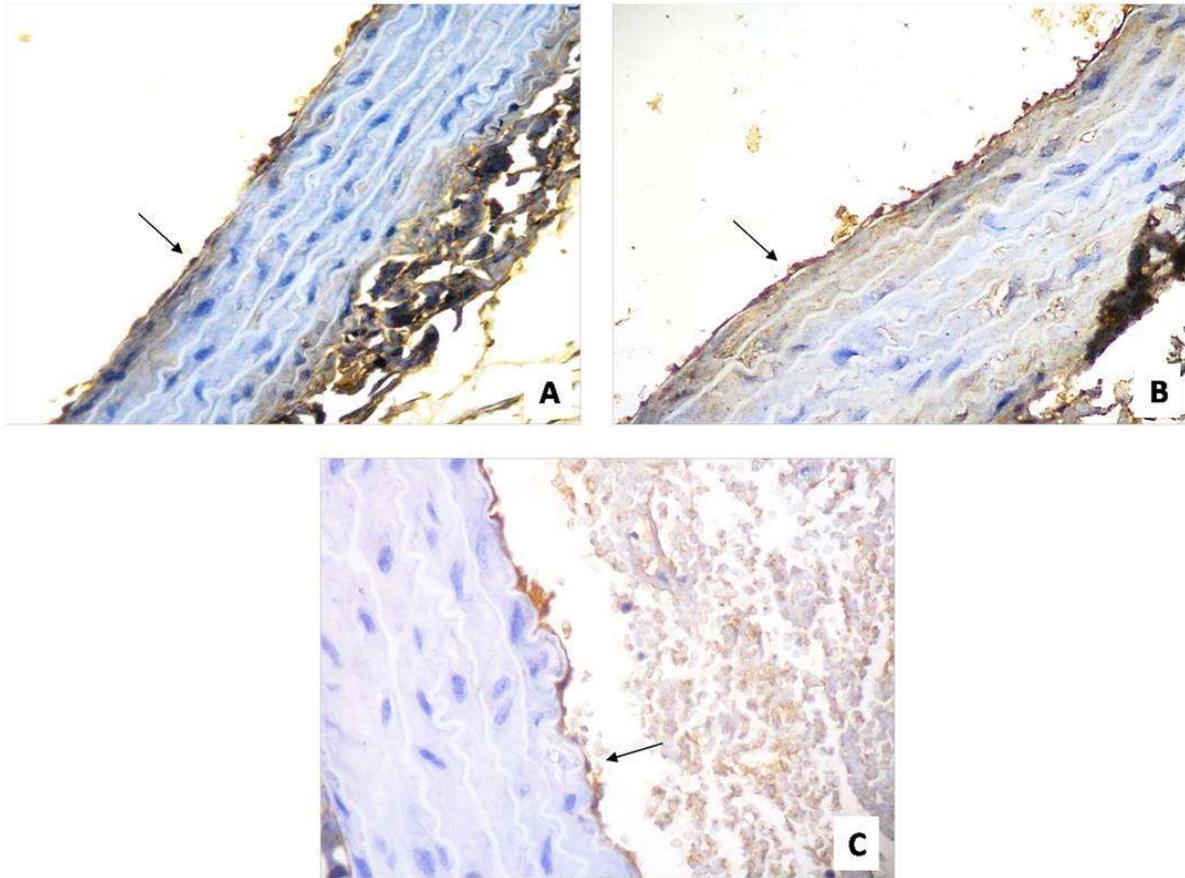
A significant decrease in percentage of  $\beta$ -catenin<sup>+</sup> cells obtained from vehicle-treated HFD rats' aortae was observed versus normal control. However, 8-week treatment with vitamin K<sub>2</sub> has significantly increased the percent of  $\beta$ -catenin<sup>+</sup> cells versus vehicle-treated HFD rats (**Figure 2 D and Figure 5 A-C**)



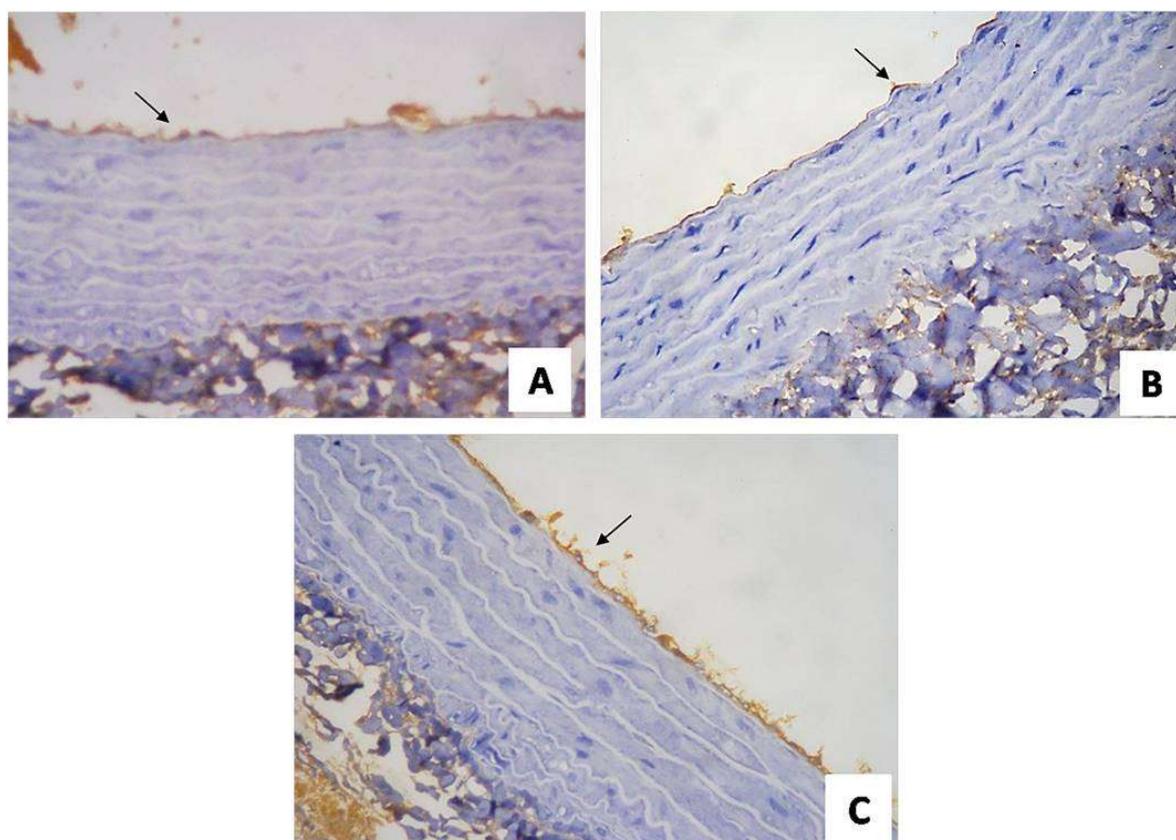
**Figure 2:** Immunohistochemical semiquantitative analysis of vitamin K<sub>2</sub> effect on HFD changes in aortic CD133<sup>+</sup> cells (A), VEGFR2<sup>+</sup> cells (B), combined CD133<sup>+</sup>/VEGFR2<sup>+</sup> cells (C), and  $\beta$ -catenin<sup>+</sup> cells (D) expression. HFD; high fat diet, K<sub>2</sub>; vitamin K<sub>2</sub>, VEGFR2; vascular endothelial growth factor receptor 2. Data are expressed as means  $\pm$  S.D. \*: significant difference versus normal control, #: significant difference versus vehicle-treated HFD.



**Figure 3: Photomicrographs of immunohistochemically stained sections of rats' aorta for CD133<sup>+</sup> cells.** Normal control rat's aorta in (A) showing dispersed CD 133<sup>+</sup> endothelial cells (↑) with brown stained nuclei, which were decreased in vehicle-treated HFD rats (B). (C) showed an increase in number of CD 133<sup>+</sup> endothelial cells in vitamin K<sub>2</sub> treated HFD rats. (×400)



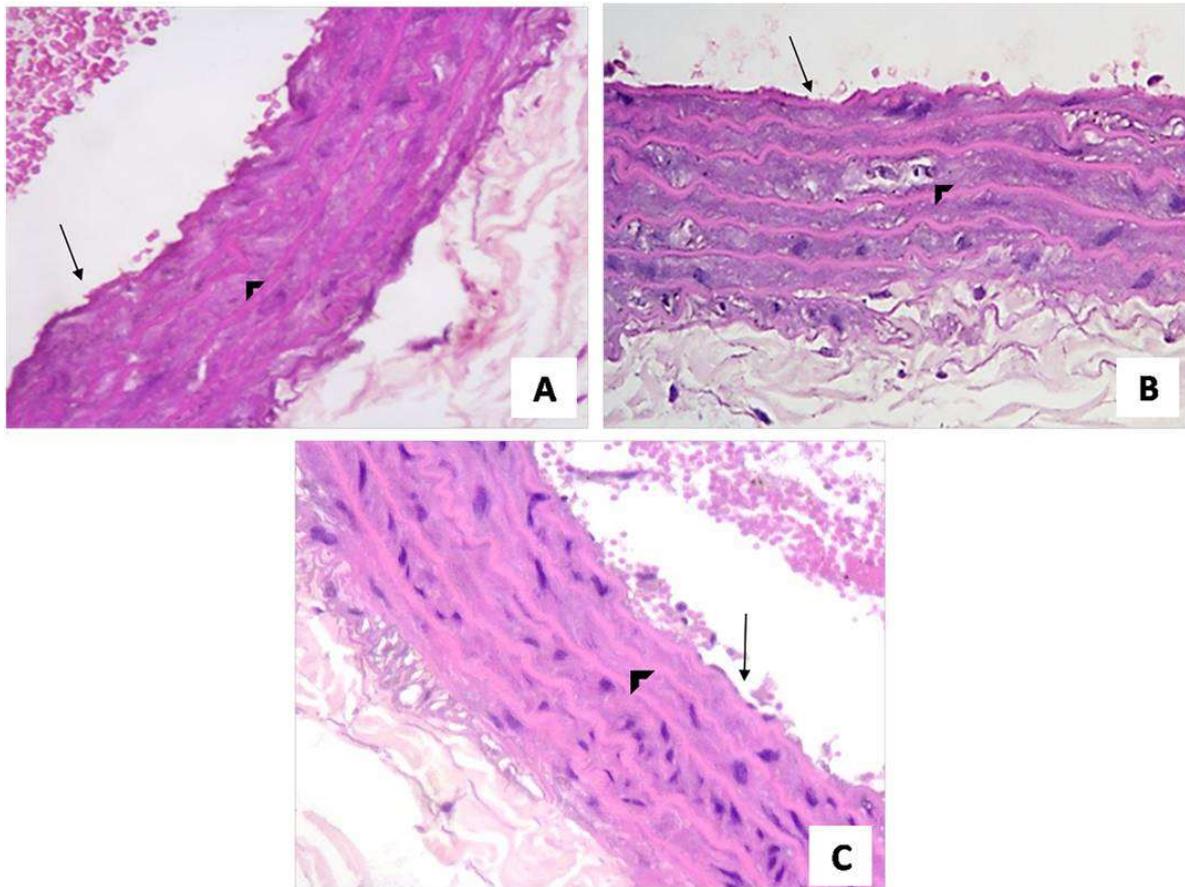
**Figure 4: Photomicrographs of immunohistochemically stained sections of rats' aorta for VEGFR2<sup>+</sup> cells.** Normal control rat's aorta in (A) showing dispersed VEGFR2<sup>+</sup> endothelial cells (↑) with brown stained nuclei, which were decreased in vehicle-treated HFD rats (B). (C) showed moderate increase in number of VEGFR2<sup>+</sup> endothelial cells in vitamin K<sub>2</sub> treated HFD rats.(×400)



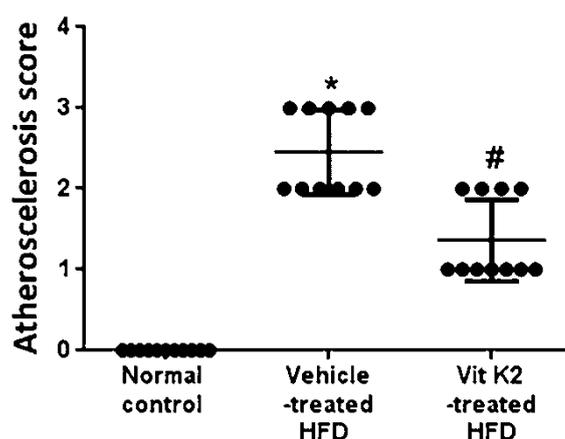
**Figure 5: Photomicrographs of immunohistochemically stained sections of rats' aorta for beta-catenin<sup>+</sup> cells.** Normal control rat's aorta in (A) showing dispersed beta-catenin<sup>+</sup> endothelial cells (↑), which were decreased in vehicle-treated HFD rats (B). (C) showed marked increase in number of beta-catenin<sup>+</sup> endothelial cells in vitamin k<sub>2</sub>treated HFD rats.(×400).

*Effect of vitamin k<sub>2</sub> on the histopathological features in HFD aortae*

Administration of HFD for 8 weeks revealed early atherosclerotic changes in comparison to normal control in H&E staining in the form of intimal irregularity with focal loss of the endothelium. The underlying tunica media appeared thin and showed fatty streaks formed of aggregates and dispersed foam cells. All these structural derangements were partly ameliorated after vitamin K<sub>2</sub> treatment (**Figure 6 A-C**) as evidenced by the semiquantitative analysis, where a significant decrease in fatty aggregates in comparison to the vehicle-treated HFD rats' aortae was noted in vitamin K<sub>2</sub> treated group (**Figure 7**).



**Figure 6:**Photomicrographs of H&E staining of aortic tissue sections, showing in (A), normal control; a layer of flattened endothelial cells in the tunica intima (arrow) and the tunica media formed of several elastic lamellae with smooth muscle fibres in between. (B), vehicle-treated HFD rats, irregularity of tunica intima with focal endothelial shedding (arrow) and the underlying tunica media shows fatty streaks formed of aggregates and dispersed foam cells (arrow head). (C), vitamin K<sub>2</sub>-treated group; minimal irregularity of the tunica intima (arrow) with fatty streaks in the underlying tunica media (arrow head). (×400).



**Figure 7: Atherosclerosis score.** Data are expressed as medians  $\pm$  interquartile range. \*: significant difference versus normal control, #: significant difference versus vehicle-treated HFD. HFD; high fat diet, k<sub>2</sub>; vitamin k<sub>2</sub>.

## Discussion

Cardiovascular morbidity and mortality are still rising despite the protection conferred by multiple therapies targeting major cardiovascular risk factors, which denotes the insufficient vascular repair process. An essential facet of this repair process is the EPCs signalling pathway in view of the cardiovascular risks as dyslipidaemia.

In the current study, administration of HFD for 8-weeks retrieved worsening in serum lipid profile. Instantaneously, the expression of composite surface markers of EPCs was reduced in association with endothelial NOX4 and  $\beta$ -catenin under-expression. These changes shared into vascular functional and structural damage, being evidenced by the altered histopathological features of the aortae and the elevated atherosclerotic score.

In context of dyslipidaemia, the induced inflammatory reactions could play a role in changing EPCs microenvironment and inducing apoptosis, thus impairing their ability to contribute into vascular repair.(21) The endothelial inflammatory reactions induced by dyslipidaemia is essentially vasculo-protective in nature but if not maintained under strict homeostatic control, it will perpetuate a harmful redox imbalance that will eventually impact the EPCs differentiation and life span inducing an endothelial dysfunction.(22)

One of the major factors involved in the homeostatic redox balance is the NOX4. Indeed, in the present study, a down regulation of NOX4 expression was observed that was associated with a reduction in EPCs numbers and vascular structural damage. This could emphasize on the reported NOX4 important role in vascular protection that involves the activation of the stress responsive genes within the EPCs.(23, 24)

Being a master regulator of growth control pathway,  $\beta$ -catenin activation permits its interaction with relevant transcription factors, endorsing endothelial cells' survival and EPCs' proliferation.(25)The observed  $\beta$ -catenin downregulation by dyslipidaemia could support the observed reduction in EPCs numbers, though its mechanism is not clear. Studies reported a role of Dickkopf protein, an inhibitor of Wnt signalling pathway induced by dyslipidaemia, in  $\beta$ -catenin suppression within the early atherosclerotic lesions.(26)Moreover, NOX4 downregulation with excess ROS accumulation can alter the downstream signalling of  $\beta$ -catenin diverting its positive contribution to EPCs differentiation into a pathological fate.(27)Consequently, the current findings could deduce how the altered EPCs' signalling induced by dyslipidaemia could induce an ED. This necessitates the emergence of drugs promoting the EPCs regenerative capacity.

In order to assess vitamin K<sub>2</sub> ability to promote EPCs repair, vitamin K<sub>2</sub> was given for HFD fed rats for 8 weeks. It exhibited a vascular protection against the injurious effect of HFD-induced dyslipidaemia. Vitamin K<sub>2</sub> increased EPCs' number and ameliorated vascular structural abnormalities with an increase in NOX4 and  $\beta$ -catenin expression.

In this context, vitamin K<sub>2</sub> induced increase in NOX4 expression that was associated with an anti-atherogenic potential and a vascular structural improvement emphasized on the previously reported NOX4 endothelial protective action.(28)Besides increasing NOX4 expression, vitamin K<sub>2</sub> is reported to possess an antioxidant potential. It was recently demonstrated that vitamin K<sub>2</sub> triggers the SIRT1 signaling pathway, resulting in increasing mitochondrial antioxidant superoxide dismutase deacetylation and decreasing mitochondrial ROS production.(29)

Likewise, the observed increase in  $\beta$ -catenin expression by vitamin K<sub>2</sub> could justify its induced increase in EPCs count. This could be in part achieved via activation of SIRT-1, thus favoring the involvement of  $\beta$ -catenin signalling in an EPCs reparative potential.(30)Moreover, the reported anti-inflammatory and antioxidant potentials of vitamin K<sub>2</sub> could add to its EPCs beneficial effect. Besides its ability to downregulate several inflammatory genes,(31)its antioxidant potentials entail an ability to mediate cellular redox homeostasis. The latter is mediated via restoring mitochondrial dysfunction, aerobic glycolysis, and oxidative phosphorylation.(23)Also some of vitamin K redox-cycle enzymes; as vitamin K-oxidoreductase, do possess a free radical-scavenging potentials that can regulate NOX activity (32) and inhibit lipoxygenase; thus halting the cascade of oxidant and pro-inflammatory mediators.(33)

Moreover, vitamin K<sub>2</sub> can also act in a hormone-like manner fostering interactions between key microRNAs, transcription factors, sirtuins, and histone deacetylases that can all share in tempting cell fate.(34) This could explain the observed improvement in EPCs number and upregulation of NOX4 and  $\beta$ -catenin by vitamin K<sub>2</sub>. This is in line with previous study,(35) which recorded an improvement in nitric oxide-dependent endothelial function in ApoE/LDLR<sup>-/-</sup> mice by Vitamin K<sub>2</sub> -MK-7.

On the other hand, vitamin K<sub>2</sub> improved the atherogenic profile of HFD fed rats. It thrived to reduce both the atherogenic index and the atherosclerotic score. This could be attributed to the reduction in LDL-C. This is consistent with previous studies, confirming variable antihyperlipidemic and antiatherogenic actions of vitamin K<sub>2</sub> that can eventually restrain the atherosclerotic progression.(36)

In agreement with the current work, vitamin k<sub>2</sub> was stated to decrease total cholesterol and preventing its vascular deposition as esters (36, 37) rather than on an ability to modulate HDL-C, which was lacking in this work.

Nevertheless, one cannot neglect the reported vitamin k<sub>2</sub> vasoprotective potential that pillars on its ability to promote post-translational carboxylation of matrix Gla-protein, a major endogenous calcification inhibitor.(38)This was found not only to inhibit vascular deposition of calcium crystals, but to further inhibit cholesterol deposition during the process of atherogenesis.(39)Overall, these raised vascular protective potential mechanisms of vitamin k<sub>2</sub> supplement were translated into an evident vascular structural improvement that further corroborate the contribution of the currently observed increase in EPCs number into an observed repair.

In conclusion, vitamin k<sub>2</sub> supplement was effective in preventing the injurious insults of the induced dyslipidaemia and inhibiting the progression of atherosclerosis. This is thought to be due to a possible interplay between vascular Wnt/ $\beta$ -catenin signalling and NOX4 expression that thrived to inducing EPCs repair signalling and increasing their number in addition to improving vascular structural integrity.

#### **Declaration of interest**

None.

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